



# IJSRM

INTERNATIONAL JOURNAL OF SCIENCE AND RESEARCH METHODOLOGY

An Official Publication of Human Journals



Human Journals

Research Article

July 2022 Vol.:22, Issue:1

© All rights are reserved by Abrahão Alves de Oliveira Filho et al.

## Evaluation of the Anti-Adherent Potential of the Essential Oil of *Matricaria chamomilla* L. (Asteraceae) Against Food-Contaminating Pathogenic Strains



João Henrique Anizio de Farias<sup>1</sup>, Lucas de Brito Silva<sup>2</sup>, Jucihermes de Almeida Mariano<sup>3</sup>, Maria Alice Araújo de Medeiros<sup>4</sup>, Millena de Souza Alves<sup>5</sup>, Bernadete Santos<sup>6</sup>, Gean Carlos Xavier Campos<sup>7</sup>, Rosália Severo de Medeiros<sup>8</sup>, Raline Mendonça dos Anjos<sup>9</sup>, Maria das Graças Veloso Marinho<sup>10</sup>, Marcos Antônio Nobrega de Sousa<sup>11</sup>, Luciano de Brito Junior<sup>12</sup>, Veneziano Guedes de Sousa Rêgo<sup>13</sup>, Abrahão Alves de Oliveira Filho\*<sup>14</sup>

<sup>1\*, 2, 3, 7</sup> Undergraduate student at the Federal University of Campina Grande - UFCG, Brazil <sup>4,5,6</sup> Master's student at the Federal University of Campina Grande - UFCG, Brazil <sup>8,9,10,11,12,13,14</sup> Professor at the Federal University of Campina Grande - UFCG, Brazil

**Submitted:** 20 June 2022

**Accepted:** 25 June 2022

**Published:** 30 July 2022

**Keywords:** Food, Health, Phytotherapy, Microbiology.

### ABSTRACT

Foodborne illness is one of the world's most frequent public health problems. From this perspective, the present study's objective is to evaluate the antibacterial and anti-adherent activity of the essential oil against strains of *Escherichia coli*, *Enterobacter cloacae*, *Klebsiella pneumoniae*, and *Staphylococcus saprophyticus*. For the assays, the slant-tube technique was used to determine the Minimum Inhibitory Adherence Concentration (MIC) to glass in the presence of 5% sucrose. The experiments were performed in duplicate. From the results obtained regarding the anti-adherent activity, it was found that the essential oil was able to inhibit the adherence of *Staphylococcus saprophyticus* at a ratio of 1:2, while the chlorhexidine digluconate 0.12% prevented adherence at a ratio of 1:8, possessing greater anti-adherent activity against *S. saprophyticus* than the tested oil. However, from the *in vitro* assay with *E. coli*, it was observed that the essential oil showed greater anti-adherent activity against this pathogen when compared to 0.12% chlorhexidine digluconate, which inhibited in the proportion of 1:2, while the essential oil of *M. chamomilla* inhibited in the proportion of 1:4. In the test performed with *Klebsiella pneumoniae*, it was observed that Oe inhibited biofilm formation at a 1:1 ratio, and no inhibition was observed with the use of chlorhexidine. Regarding *Enterobacter cloacae*, it was found that both *M. chamomilla* essential oil and chlorhexidine digluconate were not able to inhibit biofilm formation at the concentrations tested. In summary, the essential oil demonstrated an anti-adherent potential against foodborne strains of *Staphylococcus saprophyticus*, *Escherichia coli*, and *Klebsiella pneumoniae*.



HUMAN JOURNALS

[www.ijssrm.humanjournals.com](http://www.ijssrm.humanjournals.com)

## INTRODUCTION

Foodborne diseases (FOBD) are one of the most frequent public health problems in the world. They are caused by the ingestion of water and food contaminated by etiological agents, mainly pathogenic microorganisms. According to the UN, every year, millions of people worldwide fall ill and many die after ingesting contaminated food. In the period between 2000 and 2019, 14,030 outbreaks of ATD were reported in Brazil, most of them resulting from infections caused by bacteria and their toxins (Brasil, 2019).

Foods, in general, are rich sources of nutrients, which when in conjunction with intrinsic factors, such as pH and humidity, favor bacterial proliferation, and proliferation can cause both health and economic losses for industries due to deterioration (Flores; Melo, 2015). There are more than 250 types of ATD in the world, and most of them are infections caused by bacteria and their toxins (Brasil, 2019). Such factors, along with consumer demand for natural products and microbial resistance to conventional drugs (Abad *et al.*, 2007), reinforce the need for studies regarding the *in vitro* antimicrobial activity of essential oils and their contribution to food preservation.

Among the pathogenic bacteria constantly identified in outbreaks involving foodborne diseases, *Staphylococcus* spp. and pathogens of the Enterobacteriaceae family, such as *Escherichia coli*, *Enterobacter* spp, and *Klebsiella pneumoniae* that although they integrate the normal human microbiota, the external addition of these microorganisms, usually causes disharmony in the organism, resulting in infectious pictures (Feitosa *et al.*, 2017; Melo *et al.*, 2018; Pereira *et al.*, 2017; Silva *et al.*, 2018). Both *Staphylococcus* spp. and *E. coli* are addressed in ANVISA's (2013) Manual of Basic Procedures in Clinical Microbiology for the Control of Healthcare-Related Infection, either because they can cause severe infections and/or present resistance to routinely used antimicrobials, as is the case of *Staphylococcus saprophyticus*, which is resistant to novobiocin.

The impact of antibiotic resistance (AR) is growing to dangerously high levels in all parts of the world (WHO, 2018). According to the Pan American Health Organization (PAHO, 2018), RA is currently one of the biggest global threats to health, food safety, and development. In addition, several studies have reported that the ability of these bacteria to associate in the form of biofilm,

either on physical or biological surfaces, is a mechanism that contributes to increased resistance to antimicrobials/sanitizers used in clinical routines and industries (Pasternak, 2009; Rosa *et al.*, 2017; Scherrer; Marcon, 2016).

The growth of biofilms in food preparation environments can facilitate food contamination and hinder the sanitation process. According to Nesse *et al.* (2014) facilities where there is food processing, the biofilm acts not only as a reservoir of potential pathogens that can favor cross-contamination between products but also as an environment for the dissemination of virulence genes, resistance, and transmissibility of etiological agents that can cause diseases to consumers.

Given the growing incidence of pathogens that are multi-resistant to conventional drugs, the importance of the search for new alternatives is emphasized. The use of natural products obtained from medicinal plants has become a viable possibility in the fight against these microorganisms, given the pharmacological properties already registered, such as antimicrobial, anti-adherent, antifungal, antioxidant, and among others (Aquino *et al.*, 2017; Costa *et al.*, 2017; Santos *et al.*, 2015). In this sense, due to the broad biological activity presented, these plants could become a possible source for the synthesis of new drugs.

*Matricaria chamomilla* L. is an herb, popularly known as chamomile and widely spread as a medicinal plant, belonging to the Asteraceae Family. Species belonging to this family are found in practically every environment in the world. It is a family that stands out for its high economic, ornamental, and medicinal importance, since it includes not only various food and ornamental species (e.g. sunflower) but also species of medicinal value, such as *M. chamomilla* (Judd, *et al* 2009).

Chamomile is a plant with roots in traditional European medicine, and today it is officially included in the pharmacopeias in several countries. The spread of its use has probably occurred due to the presence of several active ingredients, such as alpha-bisabolol, flavonoids, apigenin, and other substances of pharmacological character (Sharifi-Rad *et al.*, 2018). Studies have shown that these constituents of chamomile essential oil have several properties, such as antimicrobial activity provided by bisabolol and flavonoids, and anxiolytic and sedative properties provided by apigenin (Madrigal-Santillán *et al.*, 2014).

Under such a perspective, the present work aimed to evaluate the anti-adherent activity of *Matricaria chamomilla* essential oil on the cell viability of pathogenic food contaminating bacteria, such as *Escherichia coli*, *Staphylococcus saprophyticus*, *Enterobacter cloacae*, and *Klebsiella pneumoniae*.

## **MATERIALS AND METHODS**

### ***In vitro* tests**

#### **Study site**

The laboratory tests were performed at the Microbiology Laboratory of the Federal University of Campina Grande, Center for Health and Rural Technology, in the city of Patos, in the sertão region of Paraíba.

#### **Test substance**

The essential oil of *Matricaria chamomilla* L. was purchased from the company QUINARÍ® (Ponta Grossa - PR). For the pharmacological assays, the substance was solubilized in DMSO and diluted in distilled water. The concentration of DMSO (dimethylsulfoxide) used was less than 0.1% v/v. The project followed the rules of CGEN - Genetic Heritage Management Council, registered in the SISGEN platform under protocol number A3DD7F.

#### **Bacterial Species and Culture Media**

In the present study, for each experiment, we used one strain of *Escherichia coli* EC46, one strain of *Staphylococcus saprophyticus* SA45, one strain of *Enterobacter cloacae* ECI41, and one strain of *Klebsiella pneumoniae* KP104. All are clinical strains, belonging to the Microbiology Laboratory of the Academic Unit of Biological Sciences, and were isolated from food contaminated with the respective bacterial species. The strains were maintained on Muller-Hinton (MH) agar at 4°C. For the experiment, together, the strains were obtained from overnight cultures in MH at 37°C and diluted in sterile saline solution to obtain the final concentration of approximately  $1.5 \times 10^8$  colony forming units per mL (CFU/mL), adjusted by turbidity comparing with the 0.5 tubes of the McFarland scale (Bona *et al*, 2014).

### **Determination of the Minimum Inhibitory Adherence Concentration (MIC)**

The Minimum Inhibitory Adherence Concentration (MIC) of the essential oil was determined in the presence of 5% sucrose, according to Albuquerque *et al.* (2009) with modifications, using corresponding concentrations of the pure essential oil up to dilution 1:1024. Both *E. coli*, *E. cloacae*, *K. pneumonia*, and *S. saprophyticus* strains were grown at 37°C in Mueller Hinton broth (DIFCO, Michigan, USA), then 0.9 ml of the subculture was dispensed into test tubes, and then 0.1 ml of the solution corresponding to the oil dilutions was added. Incubation was done at 37°C for 24 hours with tubes tilted at 30°. The reading was done by visual observation of the adherence of the bacteria to the walls of the tube, after shaking the tube and/or adding fuchsin dye to the tube wall. The assay was performed in duplicate. The same procedure was performed for the positive control, 0.12% chlorhexidine digluconate (Periogard®, Colgate Palmolive Company, New York, USA). CIMA was considered the lowest concentration of the agent in contact with sucrose which prevented the bacteria from adhering to the walls of the glass tube.

### **RESULTS AND DISCUSSION**

From the present study, the Minimum Inhibitory Adherence Concentration (MIC) of the essential oil of *M. chamomilla* L., against *Escherichia coli*, *Klebsiella pneumoniae*, and *Staphylococcus saprophyticus* strains, according to the different concentrations suggested in the methodology, was evidenced.

From the study performed, it was observed that the essential oil was able to inhibit the adherence of the *Escherichia coli* strain at a ratio of 1:4, while the 0.12% chlorhexidine digluconate prevented adherence at a ratio of 1:2, demonstrating that the essential oil showed greater anti-adherent activity against *E.coli* than the chlorhexidine digluconate, as can be seen in table 1.

**Table No. 1: Minimum Inhibitory Adherence Concentration (MIC) in µg/mL of *Matricaria chamomilla* L. essential oil and 0.12% chlorhexidine digluconate, against food-contaminating *Escherichia coli*.**

Substance	Strain	<i>Escherichia coli</i> EC46
Essential oil and <i>Matricaria chamomilla</i> L.		1:4
Chlorhexidine digluconate and 0,12%		1:2

**Fonte:** Autores, 2021.

Chlorhexidine digluconate is a very powerful synthetic antiseptic, and precisely due to its high efficacy, it is usually the reference standard compound for studies that aim to elucidate the potential of possible antimicrobial substances (Gunsolley, 2006; Kluk *et al.*, 2016).

In line with the present study, in an assay performed by Albuquerque *et al* (2010), it was also possible to observe an anti-adherent effect of chamomile extract, on dental biofilm microorganisms, such as the bacterial species *Streptococcus mutants* (ATCC 25175), *Streptococcus sanguinis* (ATCC 10557), and *Lactobacillus casei*, thus proving the anti-adherent potential of the plant species under study.

The anti-adherent potential of *M. chamomilla* L. essential oil against *E. coli* is promising, since chlorhexidine digluconate, which is the main chemical agent used as an adjuvant for biofilm control, may present adverse effects and was shown to be less effective than the essential oil tested.

Lima *et al.* (2021), when evaluating the anti-adherent potential of *Lavandula hybrida* Grosso oil against *Escherichia coli* strains, also presented results that corroborate with the present research, in which they elucidated the pathogen's sensitivity to this natural agent, which was more effective than chlorhexidine digluconate, as reported in this research on the essential oil of *Matricaria chamomilla*.



In a study by Santos *et al.* (2005), the presence of *K. pneumoniae* was found in 80 percent of the samples of enteral foods prepared in a homemade way. In this context, Straccialano *et al.* (2016) isolated samples of salad in nature, sold in restaurants and fast foods, not only strains of *K. pneumonia* but also *Enterobacter* sp., reinforcing the relationship between these pathogens as foodborne diseases.

According to Arnold *et al.* (2012), by synthesizing the enzyme known as *Klebsiella pneumoniae* carbapenemase (KPC), which is responsible for inactivating a large number of antimicrobial drugs, the transmissibility of strains of *K. pneumonia* through food may infer in pathologies difficult to treat through traditional antimicrobial agents, making it necessary to search for alternative therapies. In this sense, it reinforces the potential of natural products as anti-adherent agents, and as an alternative in the control and prevention of infections and biofilm formation by *Klebsiella pneumoniae*.

Moreover, from the present study, we also observed greater anti-adherent action of *M. chamomilla* essential oil against *Klebsiella pneumoniae* strain, when compared with chlorhexidine digluconate, since the 0.12% chlorhexidine digluconate could not inhibit biofilm formation on the wall of the glass tube in the concentrations tested, while the essential oil of *M. chamomilla* inhibited at a ratio of 1:1, as shown in Table 2.

**Table No. 2: Minimum Inhibitory Adherence Concentration (MICC) in µg/mL of Matricaria chamomilla L. essential oil and 0.12% chlorhexidine digluconate against Klebsiella pneumoniae, a food contaminant.**

Substance Strain	<i>Klebsiella pneumoniae</i> KP104
Essential oil and <i>Matricaria chamomilla</i> L.	1:1
Chlorhexidine digluconate and 0,12%	-

Source: Authors, 2022.

In line with this, when evaluating the anti-adherent potential of *Lavandula híbrida* Grosso essential oil against strains of *Klebsiella pneumoniae*, Souza et al. (2019) evidenced in their research that Lavandula oil was also able to inhibit the adhesion of the microorganism to the glass tube wall.

*Staphylococcus saprophyticus* is a bacterium that can be transmitted by ready-to-eat foods, as is the case of *S. saprophyticus* isolated from food, and it is generally resistant to antibiotics used in clinical routine, as evidenced in the work of Freitas et al. (2019), in which they identified a prevalence of *S. saprophyticus* in a food matrix in 28% of the samples studied, with some isolates being multidrug-resistant. Furthermore, there are reports of *S. saprophyticus* strains in eggshells exposed in popular trade, as well as in poultry meat and sausages such as salami ( Barcelos et al.,2017; Charmpi et al., 2020; Sommers et al. 2017).

In the present study, it was observed that *M. chamomilla* essential oil was able to inhibit the adherence of *Staphylococcus saprophyticus* at a ratio of 1:2, while 0.12% chlorhexidine digluconate prevented adherence at a ratio of 1:8, possessing greater anti-adherent activity against *Staphylococcus saprophyticus* than the tested essential oil, evidenced in Table 3.

**Table No. 3: Minimum Inhibitory Adherence Concentration (MICC) in µg/mL of Matricaria chamomilla L. essential oil and 0.12% chlorhexidine digluconate, against food-contaminating Staphylococcus saprophyticus**

Substance Strain	<i>Staphylococcus saprophyticus</i> SA45
Essential oil and <i>Matricaria chamomilla</i> L.	1:2
Chlorhexidine digluconate and 0,12%	1:8

Source: Authors, 2021.



In addition to the anti-adherent potential of the essential oil of *Matricaria chamomilla*, L. against *S. saprophyticus* evidenced in the present work, aljanaby (2018) reported in his research about the *in vitro* antibacterial activity of the aqueous extract of *Matricaria chamomilla*, the antibacterial activity of the extract against strains of *S. saprophyticus* isolated from the urinary tract of pregnant women with infection. Besides *M. chamomilla* L. other vegetables, such as the species *Coriandrum sativum*, have already shown antibacterial action in work performed by Zangeneh *et al.* (2018), on *Staphylococcus saprophyticus*. In this context, it is undeniable that the use of natural products is very promising in the fight against bacterial adherence and growth, as shown in the literature.

Another important microorganism with pathogenic potential transmitted through food is *Enterobacter cloacae*, having already been isolated from foods such as artisan cheeses, raw bovine milk, and green juices, as well as on surfaces of commercial food establishments (Alves *et al.*, 2015; Braga *et al.*, 2020; Duarte *et al.*, 2020; Lázaro *et al.*, 1999). According to Mezzatesta, Gona & Stefani (2012), *E. cloacae* can be responsible for various bacteremia of the lower respiratory tract, as well as endocarditis, and other pathologies. A high frequency of resistance of the bacterium to amoxicillin, ampicillin, cefoxitin, first-generation and broad-spectrum cephalosporins was reported in a study by Davin-Regli (2015), thus reinforcing the need to search for alternative therapies to treat diseases caused by this microorganism.

Based on the proposed methodology, it was not possible to evidence anti-adherent activity of both *M. chamomilla* essential oil and 0.12% chlorhexidine digluconate for the *Enterobacter cloacae* strain ECI41 food contaminant, as shown in Table 4.

**Table No. 4: Minimum Inhibitory Adherence Concentration (MICC) in µg/mL of *Matricaria chamomilla* L. essential oil and 0.12% chlorhexidine digluconate against *Enterobacter cloacae*, a food contaminant.**

Substance / Strain	<i>Enterobacter cloacae</i> ECI41
Essential oil and <i>Matricaria chamomilla</i> L.	-
Chlorhexidine digluconate 0,12%	-

**Source:** Authors, 2022.

Pequeno *et al.* (2018) when testing the anti-adherent effect of *Matricaria chamomilla* hydroalcoholic extract in reducing biofilm of *C. Albicans* and *Enterobacter cloacae in vitro*, observed that there was a significant reduction of *Enterobacter cloacae*, differing with the results exposed in this study, since the essential oil of *M. chamomilla* essential oil was unable to inhibit the formation of *Enterobacter cloacae* biofilm on the glass tube wall, as was 0.12% cloredixindigluconate.

Among the diverse range of natural products employed in alternative therapies (e.g. aromatherapy), essential oils are described as products with great potential for therapeutic, pharmacological, and food preservative purposes (Santos *et al.*, 2018). Such potential is verified in a study conducted by Santurio *et al* (2007), who, when evaluating the antimicrobial activity of the essential oils of oregano and thyme, common herbs in Brazilian cuisine, obtained expressive antibacterial results against serovars of enteric Salmonella, a microorganism widely associated with STDs.

Studies such as that of Tavares *et al.* (2014), have already shown that it is feasible to use an edible topping plus essential oils, such as *Origanum vulgare* (Oregano), to promote an increase in the shelf life of the food, without altering its sensory characteristics, due to the antibacterial potential of the essential oil.

Santos *et al.* (2018), prepared an Edible Coating based on alginate containing OEs of *Cinnamomum cassia* and *Myristica fragrans*, and it was verified an extension in the shelf life of apples that were coated by this RC, for about 15 days, has resulted in the decrease in the browning index and reduction of 3 log cycles of *E. coli*, a pathogen associated with fecal contamination and consequently the STD's, thus demonstrating the inhibitory potential of these natural products against microbial growth that causes deterioration/contamination of food.

In this context, besides the anti-adherent potential against food contaminating bacteria evidenced in this research, the hydroalcoholic extract obtained from the plant species under study, also showed antimicrobial activity against other microorganisms, as exposed in research conducted by Haluch *et al.* (2020), which showed positive results against strains of *Candida albicans*, and is in line with the analysis performed by Sima *et al.* (2020), about the active peptide MCh-AMP1 of chamomile, in which the results showed cell death of *Candida albicans* ATCC 10231.

Thus, studies such as those mentioned above, show the antimicrobial potential of essential oils in combating microorganisms that contaminate food, which can generate both economic losses and the health of consumers, Besides ethnopharmacological research also report the popular use of medicinal plants by traditional communities in fighting diseases, as is the case of herbs such as thyme, oregano and also chamomile, reinforcing the need for studies such as those conducted in this work, for better elucidation of the antibacterial potential of chamomile essential oil (Messias *et al.* 2015; Nunes, 2019).

## CONCLUSION

In summary, through the experiments performed in this study, according to the values obtained for the Minimum Inhibitory Adherence Concentration, it is concluded that the essential oil presents an anti-adherent potential against three of the four bacterial strains used in this study. However, further studies are needed to clarify the potential of *Matricaria chamomilla* L. essential oil anti-adherent against the cell viability of other bacteria of medical importance that contaminate food and are related to outbreaks of ATD.

## REFERENCES

1. ALBUQUERQUE ACL, PEREIRA MSV, PEREIRA JV, PEREIRA LF, SILVA DF, MACEDO-COSTA, M R (Com modificações). **Efeito antiaderente do extrato da *Matricaria recutita* Linn. Sobre microrganismos do biofilme dental.** Ver Odontol UNESP, Araraquara, 2010.
2. ALJANABY, AAJ. **In vitro antibacterial activity of aqueous extracts of *Matricaria chomomilla* flowers against pathogenic bacteria isolated from pregnant women with urinary tract infection.** Biomedical Research 2018; 29 (11): 2395-2400).
3. ALVES TS, SIQUEIRA AK, FERRAZ MMG, LEITE DS. **Identificação e perfil de sensibilidade de *Enterobacter* spp. isolados de leite bovino cru.** Vet. e Zootec. mar.; 22(1): 114-122. 2015.
4. AQUINO VVF, COSTA JGM, ANGÉLICOEC, MEDEIROS RS, ARAÚJO MF, RODRIGUESOG. **Metabólitos Secundários e ação antioxidante de *Croton heliotropiifolius* e *Croton blanchetianus*.** Acta Brasiliensis 1(3):7-10. 2017.
5. ARNOLD RS, KERRI AT, SAARIKA S, MICHAEL PJ, KRISTIE, J, MORGAN DJ. **The emergence of *Klebsiella pneumoniae* Carbapenemase (KPC)- Producing Bacteria.** South Med J. Author manuscript; available in PMC 2012 January.
6. BARCELOSD, VITOR MC, MUNHOZ AM. **Identificação bacteriana em isolados de cascas de ovos expostos em comércio popular da cidade de Guarulhos.** Revista saúde UNG. v. 11, n.1-2, 2017 ISSN 1982-3282.
7. BONA, EAM, PINTO FGS, FRUET, TK, JORGE, TCM, MOURA, AC. **Comparação de métodos para avaliação da atividade antimicrobiana e determinação da concentração inibitória mínima (cim) de extratos vegetais aquosos e etanólicos.** Arq. Inst. Biol., São Paulo, v.81, n.3, p. 218-225, 2014.
8. BRAGA LMP, SOUZA, VT, RODRIGUESMM, RAMOS GLP, GOMEASG. **Análise microbiológica de suco verde comercializado por uma rede varejista na cidade do rio de janeiro.** Alimentos: Ciência, Tecnologia e Meio Ambiente. v. 1, n. 12. 2020.
9. BRASIL. Agência Nacional de Vigilância Sanitária. **Microbiologia Clínica para o Controle de Infecção Relacionada à Assistência à Saúde. Módulo 6: Detecção e identificação de bactérias de importância médica.** Brasília: Anvisa, 2013. Disponível em: < [https://www20.anvisa.gov.br/segurancadopaciente/index.php/publicacoes?task=callelement&format=raw&item\\_id=317&element=f85c494b-2b32-4109-b8c1-083cca2b7db6&method=download&args\[0\]=c1d5d70d29b321c2507fb95630613187](https://www20.anvisa.gov.br/segurancadopaciente/index.php/publicacoes?task=callelement&format=raw&item_id=317&element=f85c494b-2b32-4109-b8c1-083cca2b7db6&method=download&args[0]=c1d5d70d29b321c2507fb95630613187) > Acesso em: 21 mai. 2022.
10. BRASIL. Ministério da Saúde, 2019. Situação epidemiológica - doenças transmitidas por alimentos. **Banco de dados 2000 a 2019.** Disponível em: < <https://www.saude.gov.br/images/xls/2020/May/11/Dados-Surtos-DTA-2000-a-2019.xls> >. Acesso em: 21 mai. 2022.
11. BRASIL. Ministério da Saúde. **Doenças transmitidas por alimentos: causas, sintomas, tratamento e prevenção.** 2019. Disponível em: <<http://www.saude.gov.br/saude-de-a-z/doencas-transmitidas-por-alimentos>> . Acesso em: 21 mai. 2022.
12. CARVALHO AF, *et al.* **Avaliação da atividade antibacteriana de extratos etanólicos e de ciclohexano a partir das flores de camomila (*Matricaria chamomilla* L.).** Revista Brasileira de Plantas Medicinais, v. 16, n. 3, p. 521-526, 2014.
13. CHARMPI *et al.* **Raw meat quality and salt levels affect the bacterial species diversity and community dynamics during the fermentation of pork mince.** Food Microbiology, v. 89, p. 103434, 2020.
14. COSTA OS, SOUZAEB, BRITO EHS, FONTENELLE ROS. **Atividade antimicrobiana e potencial terapêutico do gênero *Lippia sensu lato* (Verbenaceae).** Hoehnea 44(2): 158-171. 2017.
15. DAVIN-REGLI A, PAGÈS JM. ***Enterobacter aerogenes* and *Enterobacter cloacae*; are versatile bacterial pathogens confronting antibiotic treatment.** Front Microbiol. 2015.
16. DUARTE PN, SANTANA VTP, FERNANDESUA, DALMAS AD, HISTER BDC, FERRIIEB, DAMIÃO GM. **Avaliação microbiológica de queijos artesanais comercializados em Primavera do Leste-MT.** Ciência e Tecnologia dos Alimentos–Volume 6/ Organização Editora Poisson – Belo Horizonte - MG: Poisson. 2020.

17. FARIAS LF, *et al.* **Avaliação da atividade antibacteriana de extrato etanólico da *Bauhinia forficata* L.** *Diversitas Journal*, v. 13, n. 2, p. 402-411, 2018.
18. FEITOSAAC, SILVA JFM, RODRIGUES RM. ***Staphylococcus aureus* em alimentos.** - Revista Interdisciplinar Da Universidade Federal Do Tocantins, nº 4, 2017. p. 415-431.
19. FLORESAMPC, MELO CB. **Principais bactérias causadoras de doenças de origem alimentar.** *Revista Brasileira de Medicina Veterinária*. Nº 37. p. 65-72. 2015.
20. FREITASA, FERNANDES C, MARTINS C, SAAVEDRA MJ. **Prevalência de *Staphylococcus saprophyticus* numa matriz alimentar: avaliação do perfil de suscetibilidade a antibióticos.** In XII Jornadas de Bioquímica. Vila Real. 2019.
21. GUNSOLLEY JC. **A meta-analysis of six-month studies of antiplaque and antigingivitis agents.** *Journal of the American Dental Association*, v. 137, n. 12, p.1649-1657.2006.
22. HALUCHSM, *et al.* **Prospecção de novos antimicrobianos e bactericidas frente a microrganismos de interesse de saúde pública.** *Braz. J. Anim. Environ. Res.*, Curitiba, v. 3, n. 4, p. 3630-3652, out./dez. 2020.
23. JUDD WS, CAMPBELL CS, KELLOGG EA, STEVENS PF, DONOGHUE MJ. **Sistemática Vegetal: Um Enfoque Filogenético.** 3. ed. Porto Alegre: Artmed, 2009. 612p.
24. JÚNIOR FPA, LIMA BTM, ALVES BTW, MENEZES MES. **Fatores que propiciam o desenvolvimento de *Staphylococcus aureus* em alimentos e riscos atrelados a contaminação: uma breve revisão.** *Rev. Ciênc. Méd. Biol. Salvador*, v. 18, nº 1, 2019. p. 89-93.
25. LÁZARO NS, FARIAS RS, RODRIGUES DP, HOFER E. **Enterobacteriaceae oriundas de fontes humana e animal: produção de enterotoxina termoestável e nível de resistência a antimicrobianos / Enterobacteriaceae originating from of human sources and animal.** *Hig. Aliment; 13(64): 49-57, set. 1999.*
26. LEE A, LANCASTER H, GARAU J, KLUYTMANS J, MALHOTRA-KUMAR S, PESCHEL A, HARBARTH S. **Methicillin-resistant *Staphylococcus aureus*.** *Nat. Rev. Dis Primers*, 2018.
27. LIMA FO, *et al.* **Avaliação do potencial antiaderente do óleo de lavanda contra cepa de *Escherichia coli*.** *Research, Society and Development*, v. 10, n. 8, e22810817225, 2021 (CC BY 4.0) | ISSN 2525-3409 | DOI: <http://dx.doi.org/10.33448/rsd-v10i8.17225>.
28. MADRIGAL-SANTILLÁNE, MADRIGAL-BUJAJIDAR E, GONZÁLEZMT, SUMAYA-MARTÍNEZMT, GUTIÉRREZ-SALINAS J, BAUTISTA M, MORALES-GONZÁLEZ A, GARCÍA-LUNA M, AGUIAR-FAISAL JL, MORALES-GONZÁLEZ JA. **Review of natural products with hepatoprotective effects.** *World Journal of Gastroenterology*. v. 20. 2014.
29. MELO ES, AMORIM WR, PINHEIRO RE, CORREA PGN, CARVALHO SMR, SANTOS ARSS, BARROS DSB, OLIVEIRA ETAC, MENDES CA, SOUSAFV. **Doenças transmitidas por alimentos e principais agentes bacterianos envolvidos em surtos no Brasil: revisão.** *PUBVET*. v.12, n.10, a191, p. 1-9, out, 2018.
30. MESSIAS MCTB, *et al.* **Uso popular de plantas medicinais e perfil socioeconômico dos usuários: um estudo em área urbana em Ouro Preto, MG, Brasil.** *Revista Brasileira de Plantas Medicinais*, v. 17, n. 1, p. 76-104, 2015.
31. MEZZATESTA ML, GONAF, STEFANI F. ***Enterobacter cloacae* complex: clinical impact and emerging antibiotic resistance.** *Future Microbiol.* 7(7), 887–902. 2012.
32. NAÇÕES UNIDAS BRASIL. **Doenças transmissíveis pela comida matam 420 mil pessoas por ano no mundo, diz ONU.** 2019. Disponível em: <Doenças transmissíveis pela comida matam 420 mil pessoas por ano no mundo, diz ONU | ONU Brasil> . Acesso em: 21 mai. 2022.
33. NESSE LL, SEKSE C, BERG K, JOHANNESSEN KC, SOLHEIM H, VESTBY LK, URDAHLAM. **Potentially pathogenic *Escherichia coli* can form a biofilm under conditions relevant to the food production chain.** *Appl Environ Microbiol.* v.80, p.2042-2049, 2014.
34. NUNESMAC. **Conhecimento popular sobre plantas medicinais para o tratamento de sintomas climatéricos em Ouro Preto, Minas Gerais.** 2019. Monografia (Bacharelado em Farmácia). Universidade Federal de Ouro Preto, Ouro Preto, Minas Gerais, 2019.
35. ORGANIZAÇÃO MUNDIAL DA SAÚDE (OMS). **Antimicrobial resistência.** 2018. Disponível em:<<https://www.who.int/en/news-room/fact-sheets/detail/antimicrobial-resistance>>. Acesso em: 20 mai 2022.

36. ORGANIZAÇÃO PAN-AMERICANA DA SAÚDE (OPAS). **Novos dados revelam níveis elevados de resistência aos antibióticos em todo o mundo.** 2018. Disponível em: <[https://www.paho.org/bra/index.php?option=com\\_content&view=article&id=5592:novos-dados-revelam-niveis-elevados-de-resistencia-aos-antibioticos-em-todo-o-mundo&Itemid=812](https://www.paho.org/bra/index.php?option=com_content&view=article&id=5592:novos-dados-revelam-niveis-elevados-de-resistencia-aos-antibioticos-em-todo-o-mundo&Itemid=812)> . Acesso em: 20 mai 2022.
37. PEQUENO MA, SILVESTRE MR, AMÊNDOLA I, SILVA CRG, LEÃO MVP, SANTOS SSF. **Matricaria Recutita Extract (Chamomile) to Reduce *Candida Albicans* and *Enterobacter Cloacae* Biofilms: in vitro study.** RGO, Rev Gaúch Odontol. 2018;66(2):00-00. <http://dx.doi.org/10.1590/1981-863720180002000033328>.
38. PEREIRA TMF, GOIS VA, SOARES KMP, SOUZA LB, SOUSA JL. ***Staphylococcus aureus* e *Salmonella* sp. em queijos de coalho artesanais produzidos em São Rafael, Rio Grande do Norte.** Revista Verde, v.12, n.2. p. 358-361. 2017
39. QUARESMA AS. **Matricaria in Flora e Funga do Brasil.** Jardim Botânico do Rio de Janeiro. Disponível em: <<https://floradobrasil.jbrj.gov.br/FB80681>>. Acesso em: 10 jun. 2022
40. ROSA LV, KÄEFER K, CONCEIÇÃO NV, CONCEIÇÃO RCS, TIMMCD. **Formação de biofilme por *Vibrio parahaemolyticus* isolados de pescados.** Pesq. Vet. Bras. 37 (4): p. 339-345. 2017.
41. SANTOSA, NUNES T, COUTINHO TE, SILVA M. **Uso popular de espécies medicinais da família Verbenaceae no Brasil.** Revista Brasileira de Plantas Medicinai. p. 980-991. 2015.
42. SANTOS PR, VASCONCELOS ELQ, SOUZA AFL, JÚNIOR JLS, INHAMUNS AJ. **Qualidade físico-química e microbiológica de pescado congelado consumido na merenda escolar do estado do Amazonas.** PUBVET, v.12, n.5, mai., 2018. p. 1-6.
43. SANTOS SM, et al. **Edible active coatings incorporated with *Cinnamomum cassia* and *Myristica fragrans* essential oils to improve shelf-life of minimally processed apples.** Ciência Rural, v. 48, n. 12, 6 dez. 2018.
44. SANTOS WL, ANDRADE EGS, NORONHATH, VIEIRADG. **Indicador de contaminação fecal alimentar e prevenção de doenças.** Revista JRG de Estudos Acadêmicos, v. 2, n. 4, 2018. p. 150-157.
45. SANTOS, HC, ADALBERTO C, SOUZA EL, SOUSA CP. ***Klebsiella pneumoniae* como agente contaminante de dietas enterais artesanais** Hig. Aliment; 19(131):58-60, maio 2005.
46. SANTURIO JM, et al. **Atividade antimicrobiana dos óleos essenciais de orégano, tomilho e canela frente a sorovares de *Salmonella enterica* de origem avícola.** Produção Animal • Cienc. Rural 37 (3), Jun 2007.
47. SARTORATTOA, et al. **Composition and antimicrobial activity of essential oils from aromatic plants used in Brazil.** Brazilian Journal of Microbiology, v. 35, n. 4, p. 275-280, 2004.
48. SCHERRER JV, MARCON LNM. **Formação de biofilme e segurança dos alimentos em serviços de alimentação.** Rasbran – Revista Brasileira de Nutrição. São Paulo, SP, ano 7, n.2, p. 91-99. 2016.
49. SHARIFI-RADM, NAZRUK J, POLITO L, MORAIS-BRAGA MFB, ROCHAJE, COUTINHO HDM, SALEHI B, TABANELLI G, MONTANARI C, CONTRERAS MDM, YOUAF Z, STZER WN, DEEPA RV, MARTORELL M, SUREDA A, SHARIFI-RAD J. **Matricaria Genus as a source of antimicrobial agents: From farm to pharmacy and food applications.** Microbiological Research 215. 76-88 p. 2018.
50. SIMA SS, KHANI S, ESLAMIFAR A, AJDARY S, GOUDARZI M, HALABIAN R, AKABARIR, ZARE-ZARDINI H, FOOLADI AAI, AMANI J, ABYANEH MR. **The antifungal peptide MCh-AMP1 derived from *Matricaria chamomilla* inhibits *Candida Albicans* growth via inducing ROS Generation and altering fungal cell membrane permeability.** Frontiers in Microbiology. 2020. DOI: 10.3389/fmicb.2019.03150.
51. SOMMERS et al. **Inactivation of *Staphylococcus saprophyticus* in chicken meat and purge using thermal processing, high-pressure processing, gamma radiation, and ultraviolet light (254 nm).** Food Control, v. 75, p. 78–82, 2017.
52. SOUZA ERL, CRUZ JHA, FERREIRA JLS, OLIVEIRA HMBF, OLIVEIRA AAF. **Potencial antimicrobiano e antiaderente do óleo essencial de Lavandula híbrida Grosso contra cepas de *Klebsiella pneumoniae*.** Arch Health Invest 10(6) 2021.



53. STRACCIALANO FFL, PAULINO NTR, BRAGA AVU, MORELLI S, SANTOS RFS. **Qualidade microbiológica de saladas in natura servidas em restaurantes e fast foods na cidade de Campinas e região.** Hig. Aliment; 30(256/257): 123-127, maio/junho 2016.
54. TAVARES FO, *et al.* **Cobertura comestível adicionada de óleos essenciais de orégano e alecrim para uso em ricota.** Revista do Instituto de Laticínios Cândido Tostes, Juiz de Fora, v. 69, n. 4, 249-257, 2014.
55. ZANGENEH MM, ZANGENEH A, MORADIR, SHAHMOHAMMADI A. **Chemical Characterization and Antibacterial Activity of the Essential Oil of *Coriandrum sativum* Leaves in the West of Iran (Kermanshah).** *Journal of Essential Oil Bearing Plants*, 21(5), 2018, 1349–1358. doi:10.1080/0972060x.2018.15261.

