



# IJSRM

INTERNATIONAL JOURNAL OF SCIENCE AND RESEARCH METHODOLOGY

An Official Publication of Human Journals



Human Journals

Research Article

March 2022 Vol.:21, Issue:1

© All rights are reserved by Onitsha, Enebrayi N et al.

## Lipid Profile and Malondialdehyde Levels of Mortuary Attendants Exposed to Embalming Chemicals in Some Mortuaries in South-South, Nigeria



### IJSRM

INTERNATIONAL JOURNAL OF SCIENCE AND RESEARCH METHODOLOGY

An Official Publication of Human Journals



Onitsha, Enebrayi N<sup>1\*</sup>, Ugochukwu Chioma

Promise<sup>2</sup>, Ofor, Igri B.<sup>3</sup>

<sup>1\*</sup>Department of Medical Laboratory Science, Faculty of Basic Sciences, College of Health Science, Niger Delta University, Wilberforce Island, Amassoma, Bayelsa State, Nigeria.

<sup>2</sup>Department of Medical Laboratory Sciences, College of Health Sciences, Nnamdi Azikiwe University, Nnewi, Anambra State, Nigeria.

<sup>3</sup>Medical Laboratory Services, Federal Medical Centre Yenagoa, Bayelsa State, Nigeria

**Submitted:** 21 February 2022

**Accepted:** 26 February 2022

**Published:** 30 March 2022



HUMAN JOURNALS

[www.ijssrm.humanjournals.com](http://www.ijssrm.humanjournals.com)

**Keywords:** Embalming Chemicals, Formaldehyde, Lipid Profile Parameters, Malondialdehyde, and Mortuary Attendants.

### ABSTRACT

**Background:** Human exposure to embalming fluid containing formaldehyde is associated with multiple negative effects, and its exposure is known to cause oxidative stress in some vital organs like the liver, kidney, and lungs. The drought of basic data on the effect of embalming fluids on biochemical parameters, as well as safety precautions in the use of these chemicals, underscores the need for more extensive studies on embalming chemicals and their related health effects. **Aim:** This study evaluated the effect of embalming chemicals on lipid profile parameters and malondialdehyde (MDA) levels of mortuary attendants in some selected mortuaries in Bayelsa State, Nigeria. **Method:** In this work, a total of twenty-five (25) mortuary attendants, within the age range of 30-50 years, are exposed to embalming chemicals for a period of two (2) to twenty (20) years. Also, twenty-five (25) apparently healthy unexposed individuals within the same age range were recruited as controls. Blood samples were collected from the subjects and estimation of plasma lipids and malondialdehyde (MDA) levels were done using the spectrophotometric method. **Result:** The results showed that total cholesterol ( $4.67 \pm 0.56$ ) and triglyceride ( $1.91 \pm 0.86$ ) levels of the mortuary attendants were slightly higher than the control ( $4.51 \pm 0.37$  and  $1.78 \pm 0.48$  respectively), but was not significant ( $P > 0.05$ ). High-Density Lipoprotein ( $1.32 \pm 0.306$ ) showed a statistically significant ( $p < 0.05$ ) increase in the exposed subjects when compared with the control ( $0.96 \pm 0.88$ ). However, there was a significant ( $p < 0.05$ ) decrease in the level of Low-Density Lipoprotein ( $2.25 \pm 0.89$ ) of mortuary attendants compared with the unexposed subjects ( $3.45 \pm 0.99$ ). The serum malondialdehyde ( $3.27 \pm 0.530$ ) level was significantly ( $p < 0.05$ ) higher in the exposed workers than unexposed subjects ( $1.78 \pm 0.303$ ). Furthermore, there was a statistically significant ( $p < 0.05$ ) elevation in malondialdehyde ( $3.78 \pm 0.604$ ) among exposed subjects with  $> 5$  years duration compared with  $< 2$  years ( $1.84 \pm 0.42$ ). **Conclusion:** This study confirms that exposure to embalming fluid alters serum lipids and increases lipid peroxidation product malondialdehyde (MDA) in humans. This elevation is based on the prolonged duration of exposure. Thus, the use of personnel protective equipment is recommended for morticians.

## INTRODUCTION

Occupational exposure to toxic chemicals in workplaces portends a serious risk to human health [1]. Mortuary workers are exposed to toxic chemicals such as preservatives, buffers, anticoagulants, germicides, fungicides, perfuming agents, hygroscopic agents, and dyes present in embalming fluids which are known to cause several health complications [2]. Some of these complications include; respiratory disorders, genotoxicity, dermatitis, eye blindness, autoimmunity, ocular irritations, corneal clouding, leukemia, nasopharyngeal cancers, congenital malformations, and menstrual irregularities [3].

Embalming is the process of temporarily preserving human remains to prevent decomposition and prepare them for public display at funerals [4]. It involves the application of chemicals to a dead human body in order to reduce the presence and growth of the microorganisms, retard organic decomposition, and restore acceptable physical appearance [5]. Embalming chemicals are majorly composed of preservatives, sanitizing, and disinfection agents and additives used in modern embalming [6]. Some essential constituents in embalming chemicals are buffers, anticoagulants, germicides, fungicides, perfuming agents, hygroscopic agents, and additives [2]. The goals of modern-day embalming are disinfection, preservation, and restoration of the body. Embalming fluids or chemicals are administered via arterial and intramuscular injection or sprayed on the body. The general principle is that embalming fluids act to fix cellular proteins [2]. Formaldehyde fixes tissues or cells by irreversibly connecting a primary amine group in protein molecule with nearby nitrogen or DNA molecule through CH<sub>2</sub> linkage called Schiff base [6]. During the 19<sup>th</sup> and 20<sup>th</sup> centuries, arsenic-based solutions were the first widely recognized and regularly used embalming fluid, although they have since been replaced by more effective and less toxic formaldehyde [7].

The embalming fluid of today typically contains a mixture of formaldehyde, methanol, ethanol, phenol glutaraldehyde, glycerine, oil of wintergreen, eosine solution, phenoxyethanol, and other solutions [8]. Of all the chemicals used in modern-day embalming, formaldehyde has been found to have the most important exposure concern. It is absorbed via the respiratory and gastrointestinal routes [3]. It has been reported that long-term exposure to formaldehyde at concentrations exceeding the national standard exerts a variety of toxic effects on the nasopharynx, lung, brain, and skin, and also on the hematopoietic organs bone marrow (BM) and

the spleen [8]. On the basis of epidemiological studies, the International Agency for Research on Cancer (IARC) classified formaldehyde as a human leukemogenic [8], while some studies on humans and animals showed that occupational exposure to formaldehyde can disrupt hematopoietic function and lead to hematopoietic toxicity [9, 10].

Lipids are organic substances relatively insoluble in water but soluble in organic solvents and are utilized by living cells. They are structural components in cells and are involved in metabolic and hormonal pathways [11]. Measurement of serum lipids levels has been the prime index of cardiovascular disease (CVD). However, several “atherogenic indices” such as TC/HDL-C and LDL-C/HDL-C ratios are considered risk indicators with greater predictive value than isolated parameters used independently, particularly LDL-C [12]. TG/HDL-C ratio also has been reported as a significant predictor of extensive coronary heart disease [13].

Malondialdehyde (MDA) is a final lipid peroxidation product and it is the most widely used indicator of the degree of the oxidation process in body fluids [14]. Lipid peroxidation is a well-established mechanism of cellular injury in humans and is used as an indicator of oxidative stress in cells and tissues. Lipid peroxides derived from polyunsaturated fatty acids are unstable and can be decomposed to form a complex series of compounds. These include reactive carbonyl compound, which is the most abundant malondialdehyde (MDA) [15]. The measurement of MDA is widely used as an indicator of lipid peroxidation and increased levels of the peroxidation products have been associated with a variety of acute, chronic pathophysiological processes in the human as well as animal models [16].

This study was conducted to evaluate the effects of embalming chemicals on plasma lipids and malondialdehyde levels of mortuary workers in some mortuaries in South-South, Nigeria.

## **METHODS**

### **Study Population**

A total of fifty (50) subjects were recruited for the study, which comprises twenty-five (25) mortuary attendants exposed to embalming chemicals and twenty-five (25) apparently healthy unexposed subjects who served as a control group. All the fifty (50) subjects used for the study were male with an age range of 20–50 years. Subjects who work in the morgue and consented to

the study without a known medical history of any metabolic disorder were included in the study. Subjects who do not consent to the study and with a medical history of known metabolic disorder were excluded from the study. Also excluded are chronic cigarette smokers and chronic alcohol drinkers. The ethical clearance was approved by the research ethical committee of Federal Medical Centre (FMC) Yenagoa, Niger Delta University Teaching Hospital Okolobiri (NDUTH), and other private mortuaries in South-South, Nigeria.

### **Blood Sample Collection**

Seven (7) milliliters of blood sample was collected from each of the fifty (50) subjects after fulfilling the inclusion and exclusion criteria. Five (5) milliliters of the blood sample were dispensed into an Ethylene diamine tetraacetic acid (EDTA) sample tube. The remaining two (2) milliliters were dispensed into a plain dry glass tube. The blood samples were transported to the Research Laboratory of Medical Laboratory Science, Niger Delta University in a cool box containing ice bags. The samples were centrifuged at 3000rpm for 10minutes to obtain the clear serum and plasma. They were separated into separate plastic dry sample tubes and stored at -20<sup>0</sup>C and analysis were done within 48hours of sample collection. Total cholesterol (TC), High-Density Lipoprotein (HDL), and triglyceride (TRIG) were measured using reagents from Randox Diagnostic kits as specified by Randox Diagnostics (Switzerland). Serum Lipid peroxide analysis was carried out by determining the concentration of MDA formed using the method of Varshney and Kale <sup>[17]</sup>.

### **Analysis of Biochemical Parameters**

**Determination of lipid profile Parameters:** Lipid profile parameters: total cholesterol (TC), High-Density Lipoprotein Cholesterol (HDL-C), and Triglycerides (TG) were estimated by spectrophotometric methods using Randox reagent kits. Plasma Total Cholesterol concentration was measured by the enzymatic method as described by Tindler <sup>[18]</sup> and modified by Richmond, [19], and high-density lipoprotein-cholesterol (HDL-C) and low-density lipoprotein-cholesterol (LDL-C) were determined enzymatically after precipitation of other lipoproteins as described by Burstein *et al.*, <sup>[20]</sup> and Assmann *et al.*, <sup>[21]</sup> respectively, using Randox reagent kits.

**Determination of Malondialdehyde:** Serum Malondialdehyde (MDA) was measured by the method of Shah and Walker's, <sup>[22]</sup> using an auto-analyzer spectrophotometer. In this reaction, malondialdehyde conjugates with thiobarbituric acid (TBA) reagent under acidic conditions to generate a pink-colored product, and the absorbance was determined at 532 nm. Briefly, 1.0ml reagent 1 (17.5% TCA), reagent 2 (70% TCA) and reagent 3 (Thiobarbituric acid 0.6%) was added to 1.0ml of serum and mixed. The reaction mixture was incubated in a boiling bath for 15 minutes, allowed to cool, and then let to stand at room temperature for another 20 minutes. Then the tubes were centrifuged at 2000 rpm for 15 minutes and the supernatant layer was read at 534 nm. Distilled water was used for the blank. The concentration of MDA (nmol/ml) was calculated by using the following formula: Concentration of the test= Abs (test) –Abs (blank) / 1.56 x 1000000

## DATA ANALYSIS

Results obtained from the biochemical estimations were analyzed with SPSS version 23.0. Data presentations were in the form of tables. The mean and standard deviation of the mean values of lipid profile parameters of mortuary workers were compared with values of those not exposed to the embalming chemical using student t-test and ANOVA. The level of significance was set at  $P < 0.05$ .

## RESULTS

**Table 4.1** shows the Comparison of the Mean level of Serum Lipid Profile Parameters among Mortuary Workers and the Control Group. The result showed that the mean values of total cholesterol ( $4.67 \pm 0.56$ ), High-density lipoprotein ( $1.32 \pm 0.306$ ), and triglyceride ( $1.91 \pm 0.86$ ) of the mortuary workers were slightly higher than the control group ( $4.51 \pm 0.37$ ;  $0.96 \pm 0.88$  and  $1.78 \pm 0.48$  respectively). However, there was no statistically significant ( $P > 0.05$ ) difference in the mean values of total cholesterol and triglyceride, while high-density lipoprotein (HDL) shows a statistically significant ( $p < 0.05$ ) difference. The mean value of low-density lipoprotein ( $2.25 \pm 0.89$ ) shows a statistically significant ( $P < 0.05$ ) reduction in the exposed workers when compared with the control group ( $3.45 \pm 0.99$ ).

**Table 4.2** shows the comparison of the Level of Lipid peroxidation product Malondialdehyde among Mortuary Workers and Control Group. The result revealed that the mean value of malondialdehyde ( $3.27 \pm 0.530$ ) of the mortuary workers exposed to the embalming chemical is significantly ( $p < 0.05$ ) higher than the control group ( $1.78 \pm 0.303$ ).

**Table 4.3** shows the Statistical results of the effect of duration of exposure to embalming chemicals on lipid profile parameters and malondialdehyde level of mortuary workers. The result revealed that the mean values of total cholesterol, high-density lipoprotein, and triglyceride levels showed a statistically non-significant ( $p > 0.05$ ) increase across the three groups of mortuary workers, while low-density lipoprotein shows a statistically non-significant ( $p > 0.05$ ) increase across the three groups of mortuary workers when compared with the control group. However, the mean value of MDA ( $3.78 \pm 0.604$ ) showed a statistically significant ( $p < 0.05$ ) increase in the mortuary workers of  $> 5$  years exposure to embalming chemicals when compared with the control group ( $1.78 \pm 0.303$ ).

**Table 4.1: Comparison of Mean level of Serum Lipid Profile Parameters among Mortuary Workers and Control Group**

PARAMETER	CONTROL n= 25 $\bar{X} \pm SD$	MORTUARY WORKERS (n=25) $\bar{X} \pm SD$	P-VALUE	REMARK
Total CHOL	$4.51 \pm 0.37$	$4.67 \pm 0.56$	0.930	NS
HDL	$0.96 \pm 0.88$	$1.32 \pm 0.306$	0.00	S
LDL	$3.45 \pm 0.99$	$2.25 \pm 0.89$	0.002	S
TRIG	$1.78 \pm 0.48$	$1.91 \pm 0.86$	0.296	NS

**Key:** Results are expressed as Mean  $\pm$  Standard Error of Mean (SEM).  $P < 0.05$  is considered significant, S=Significant, NS=Non-significant. Total CHOL= Total Cholesterol; HDL= High Density Lipoprotein; LDL= Low Density Lipoprotein; TRIG= Triglyceride.

**Table 4.2: Comparison of the Level of Lipid peroxidation product Malondialdehyde Among Mortuary Workers and Control Group.**

PARAMETERS	CONTROL n= 20 (X ± SD)	MORTUARY WORKERS (X ± SD)	P- VALUE	REMARK
MDA (µmo/L)	1.78±0.303	3.27±0.530	0.001	S

**Keys:** Results are expressed as Mean ± Standard Error of Mean (SEM). P< 0.05 is considered significant, S=Significant, NS=Non-significant.MDA= Malondialdehyde

**Table 4.3: Effect of Duration of Exposure to Embalming Chemical on lipid profile Parameters of Mortuary Workers**

PARAMETERS	CONTROL (n = 20) (X±SD)	DURATION OF EXPOSURE			P-Value
		< 2 years (n = 3) X±SD	2-5 years (n = 10) X±SD	> 5 years (n = 7) X±SD	
T CHOL (mmo/L)	4.511±0.37	4.23±0.23	4.35±0.22	5.03±0.83	0.453
HDL (mmo/L)	0.96±0.88	1.35±0.63	1.35±0.59	1.39±0.186	0.073
LDL (mmo/L)	3.45±0.99	2.79±0.50	1.77±0.49	2.61±1.17	0.237
TRIG (mmo/L)	1.98±0.48	2.11±0.45	1.588±1.11	1.52±0.507	0.521
MDA (µmol/L)	1.78±0.303	1.84±0.42	2.08±0.53	3.78±0.604	0.010*

**Key:** Results are expressed as Mean ± Standard Error of Mean (SEM). P< 0.05 is considered significant. Total CHOL= Total Cholesterol; HDL= High Density Lipoprotein; LDL= Low Density Lipoprotein; TRIG= Triglyceride.

## DISCUSSION

Chronic exposure to chemicals like formaldehyde, phenol, methanol, ethanol, which are the ingredients of the modern embalming solution, portend an adverse effect to humans. Studies have shown the toxicity of embalment chemicals, especially formaldehyde to the physiology

and homeostasis of man, including carcinogenicity and other adverse health effects <sup>[6]</sup>. However, this study was designed to evaluate possible alterations in the serum lipid parameters and lipid peroxidation indices of mortuary attendants exposed to embalming chemicals.

The study results showed that the mean values of total cholesterol, high-density lipoprotein cholesterol, and triglycerides of the attendants were slightly higher than the control group. Though, there was no statistically significant ( $P > 0.05$ ) difference in the mean values of total cholesterol and triglycerides, while high-density lipoprotein cholesterol (HDL-C) showed a statistically significant ( $p < 0.05$ ) difference. The increase in high-density lipoprotein cholesterol (HDL-C) with constant moderate exposure to ethanol or formaldehyde is thought to be associated with an increase in the transport rates (TRs) of apoA-I and -II. This hypothesis is in agreement with works by Naudet *al.*<sup>[23]</sup> who reported that low, to moderate constant ethanol exposure, would be associated with reduced risk for coronary disease by raising HDL-cholesterol.

The mean value of low-density lipoprotein cholesterol (LDL-C) showed a statistically significant ( $P < 0.05$ ) reduction in the exposed attendants when compared with the control group. The reduction in low-density lipoprotein cholesterol (LDL-C) could be associated with genetic variations in the apolipoproteins metabolism, particularly apolipoprotein A5 polymorphism. This is consistent with a previous study by Perissinotto and his team of experts <sup>[23]</sup> who reported a reduction in low-density lipoprotein cholesterol (LDL-C) levels in formaldehyde-exposed individuals.

Formaldehyde can trigger oxidative stress by increasing the formation of Reactive Oxygen Species (ROS), and for this reason, some secondary toxic effects in cardiac cells and tissues [25]. ROS can cause oxidative damage and lipid peroxidation by interacting with biological molecules such as DNA and lipids, cum the activation of oxidases, and inhibition of scavenging protocols <sup>[26]</sup>. This present study revealed that the mean value of malondialdehyde (MDA) of the mortuary attendants understudy is significantly ( $p < 0.05$ ) higher than the control group. Somewhat authenticating the works of Olisahet *al.*<sup>[27]</sup> and Tasdemiret *al.*<sup>[28]</sup> who reported that MDA levels in the formaldehyde exposed group was significantly higher when compared with the unexposed ( $P < 0.05$ ).



Consequently, the results also revealed that the mean values of total cholesterol, high-density lipoprotein cholesterol, and triglycerides levels were statistically insignificantly ( $p>0.05$ ) increase across the three groups of mortuary attendants, while low-density lipoprotein cholesterol showed a statistically non-significant ( $p>0.05$ ) increase across the three groups of mortuary attendants when compared with the control group. However, the mean value of malondialdehyde (MDA) showed a statistically significant ( $p<0.05$ ) increase in the mortuary attendants of > 5years exposure to embalming chemicals when compared with the control group. This finding is in agreement with Odiegwu and Colleagues <sup>[6]</sup> who documented that the adverse effect of exposure to formaldehyde is dependent on the duration of exposure.

## CONCLUSION

The results revealed that Mortuary attendants exposed to the embalming chemicals (formaldehyde or ethanol) showed significant variations in serum lipid profile levels. The serum malondialdehyde (MDA) level was significantly higher in the formaldehyde/ethanol exposed attendants than in the unexposed group. This alteration is based on the prolonged duration of exposure.

## ACKNOWLEDGEMENT

I sincerely thank all the authors for contributing immensely to making this article a success. I also appreciate the management and staff of Federal Medical Centre, Niger Delta University Teaching Hospital, and other morgues staff for their support in making this works a success.

## REFERENCES

1. Nwoke KU, Ezeh NP, Adienbo OM. Some Occupations and Their Effects on Hematological Parameters of Exposed Individuals in Port Harcourt, Nigeria. *Ijstrm.Human*, 2017; Vol. 8 (2): 50-57.
2. Mayer RG. *Embalming: History, Theory, and Practice* (5th ed.). New York, NY: McGraw-Hill Medical. 2012.
3. Pedro JS. Inductive and resonance effects on the acidities of phenol, enols, and carbonyl  $\alpha$ -hydrogens. *Journal of Org Chem*, 2009; 74: 914-916.
4. Martin EA. *Oxford Medical Dictionary*. (6th edn). Oxford University Press, New York. 2003; 222.
5. Arvinder P, Singh B. *International Journal of Medical Toxicology and Legal Medicine*, 2010; 12(6):478-481.
6. Odiegwu CNC, Ude RC, Onwurah OW, Okey-Onyesolu CF, Odiegwu U. O. (2018) Assessment of Some Haematological Parameters of Mortuary Workers Exposed to Embalment Chemicals in Some Mortuaries in Anambra State-Nigeria. *Journal of Hematological Thrombotic Disease*, 2018;6: 288.
7. Curtis DR. The basis of funeral services. *History of Embalming*. New York Chemical Publishing Co. New York. 2001: 34-35.

8. IARC. Monographs on the evaluation of carcinogenic risks to humans -formaldehyde. World Health Organization, International Agency for Research on Cancer, 2012; 35.
9. Wei C, Wen H, Yuan L, McHale CM, Li H, Wang K, Yuan J, Yang X, Zhang L. Formaldehyde induces toxicity in mouse bone marrow and hematopoietic stem/progenitor cells and enhances benzene-induced adverse effects. *Arch. Toxicol.* 2017; 91, 921–933.
10. Zhang L, Steinmaus C, Eastmond DA, Xin XK, Smith MT. Formaldehyde exposure and leukemia: A new meta-analysis and potential mechanisms. *Mutat. Res./ Rev. Mutat. Res.* 2009; 681, 150–168.
11. Crook, M.A. (2006) Chapter 13: Plasma Lipids and Lipoproteins. In ‘Clinical Chemistry & Metabolic Medicine’, 7th Edition. Hodder Arnold. pp. 198-213.
12. Millán J, Pintó X, Muñoz A. “Lipoprotein ratios: physiological significance and clinical usefulness in cardiovascular prevention,” *Vascular Health and Risk Management*, 2009; 5:757–765.
13. Da-LuzProtasio, Desidério F, José RF, Pedro A L. High Ratio of Triglycerides to HDL-Cholesterol Predicts Extensive Coronary Disease. *Clinics (São Paulo, Brazil)*; 200863(4):427-32.
14. Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem. Biol. Interact.* 2006; 160: 1–40.
15. Adeyinka AA, Adeyinka OA. Serum malondialdehyde levels during menstrual cycle *African Journal of Biotechnology* Vol. 4 (11), pp. 1297-1299, November 2005.
16. Kilic E, Suleyman Yazar, Recep Saraymen, HaticeOzbilge. Serum Malondialdehyde level in patients infected with *Ascaris lumbricoides*. *World J. Gastroenterol.* 2003; 9(10): 2332 – 2334.
17. Varshney R, Kale RK. Effects of calmodulin antagonist. *Int. J. Radiat. Biol.*, 1990; 58: 733-743.
18. Tinder P. Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. *Ann Clin Biochem*, 1969; 6: 24–7.
19. Richmond N. Preparation and properties of a cholesterol oxidase from *Nocardia* sp. and its application to the enzymatic assay of total cholesterol in serum. *Clin Chem*, 1973; 19: 1350–6.
20. Burstein M, Scholnick HR, Morfin R. Rapid method for the isolation of lipoproteins from human serum by precipitation with polyanions. *Scand J Clin Lab Invest*, 1980; 40: 583–95.
21. Assmann G, Jabs HU, Kohnert U, Nolte W, SchriewerH.. LDL-cholesterol determination in blood serum following precipitation of LDL with polyvinyl sulfate. *Clin Chim Acta*, 1984; 140: 77–83.
22. Shah JK, Walker's AM. Quantitative determination of MDA. *Biochemical Biophysics. Anti-counterfeiting Trade Agreement.* 1989;11:207-11.
23. Naud LM, Bensenor IJM, Lotufo PA. Lipid profile and alcohol consumption: longitudinal study on adults' health (ELSA-BRASIL). *SMAD, Rev EletrônicaSaúde Mental ÁlcoolDrog.* 2020;16(1):1-9.
24. Perissinotto E, Buja A, Maggi S, Enzi G, Manzato E, Scafato E, Matrangelo G, Sergi G. (2010). Alcohol consumption and cardiovascular risk factors in older lifelong wine drinkers: the Italian Longitudinal Study on Aging. *Nutr Metab Cardiovasc Dis.* 2010; 20(9): 647-655.
25. Wu D, Jiang Z, Gong B, Dou Y, Song M, Song, X, Tian Y. Vitamin E reversed apoptosis of cardiomyocytes induced by exposure to high dose formaldehyde during mice pregnancy. *Int. Heart J.*, 2017; 58(5):769-77.
26. Li H, Wang J, König R, Ansari GS, Khan MF. Formaldehyde- protein conjugate-specific antibodies in rats exposed to formaldehyde. *J. Toxicol. Environ. Health A.* 2007; 70(13):1071-1075.
27. Olisah, M.C, Y.A, et al. "Oxidative Stress Markers and Liver Functions of Morticians Exposed to Formaldehyde in South-Eastern, Nigeria." *IOSR Journal of Environmental Science, Toxicology and Food Technology (IOSR-JESTFT)*, 13(3), (2020): 14-18.
28. Tasdemir, R, Esra A, Tuncay C, Fatih H, Belgin B, Hale MK, Fatma C. Eraldemir. Investigation of Possible Oxidative Damage Caused by Formaldehyde Exposure in the Rat's Heart and Aorta Tissue *International Journal of Morphology* 2021; 39(4):42-44.

1 <sup>st</sup> Author  <b>Corresponding Author</b>	Onitsha, Enebrayi Nelson <sup>1*</sup>	<b>Address:</b> Department of Medical Laboratory Science, Faculty of Basic Sciences, College of Health Science, Niger Delta University, Wilberforce Island, Amassoma, Bayelsa State, Nigeria.
2 <sup>nd</sup> Author	Ugochukwu Chioma Promise <sup>2</sup>	<b>Address:</b> Department of Medical Laboratory Sciences, College of Health Sciences, Nnamdi Azikiwe University, Nnewi, Anambra State, Nigeria.
3 <sup>rd</sup> Author	Ofor, Igri Bassey <sup>3</sup>	<b>Address:</b> Medical Laboratory Services, Federal Medical Centre Yenagoa, Bayelsa State, Nigeria

